The HOX complex neighbored by the EVX gene, as well as two other homeobox-containing genes, the GBX-class and the EN-class, are located on the same chromosomes 2 and 7 in humans

Toshikazu Matsui^{a,b}, Momoki Hirai^c, Masami Hirano^b, Yoshikazu Kurosawa^{a,*}

^aInstitute for Comprehensive Medical Science and ^bDepartment of Internal Medicine, Fujita Health University, Toyoake, Aichi 470-11, Japan

Department of Anthropology, Graduate School of Science, The University of Tokyo, Bunkyo-ku, Tokyo 113, Japan

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Two newly identified human homeobox-containing genes, GBX1 and GBX2, are closely related genes, as are members of the other homeobox genes, EN-1 and EN-2. GBX1 and EN-2 have been mapped to chromosome 7q36. The present study shows that GBX2 was mapped to chromosome 2q37. EN-1 was mapped to chromosome 2q14. Moreover, two HOX complexes neighbored by the EVX gene, HOXA and HOXD, are located at chromosome 7p15-p14 and 2q31-q37, respectively. Thus, it is possible that these homeobox genes were linked to each other on an ancestral genome and that the ancestral chromosome segment was duplicated during evolution.

Homeobox gene; Linkage group; Evolution

1. INTRODUCTION

The homeobox was first identified as a common sequence in several homeotic and segmental genes of Drosophila, such as Antp, Ubx and Ftz [1]. Around forty homeobox genes have been isolated from Drosophila to date [2]. Mouse and human homeobox genes homologous to those of *Drosophila* have also been isolated by cross-hybridization with Drosophila probes [3]. In addition, several homeobox genes, the homologues of which have not yet been isolated from Drosophila, have been identified in mouse and human by various methods. namely, hybridization with degenerate oligonucleotides that correspond to the helix 3 region of the homeodomain [4], the polymerase chain reaction (PCR) method [5], and a search for genes localized in the vicinity of the break point of chromosome translocation observed in leukemia [6]. The Antp-class genes form the largest family of homeobox genes in the human genome and they are clustered in four complexes, HOXA, HOXB, HOXC and HOXD [7]. The four HOX complexes appear to have been created by multiplication of a primordial gene cluster that corresponds to the bithorax complex and the antennapedia complex in Drosophila [8]. At the HOM and HOX loci, the clustering of genes and their

*Corresponding author. Fax: (81) (562) 93 8835.

Abbreviations: PCR, polymerase chain reaction; FISH, fluorescence in situ hybridization.

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order seem to be essential to development since these characteristics are retained in all vertebrates and arthropods. Two human homeobox genes, EVX1 and EVX2, are also linked to the HOXA and HOXD clusters on chromosomes 7 and 2, respectively [9,10] In vertebrates, in addition to HOX genes, there are many examples of the duplication or multiplication of homeobox genes as follows: EN-1 and EN-2 [11], Cdx1, Cdx2 and Cdx3 [12]; HOX7.1 and HOX8.1 [13]; Emx1 and Emx2 [14]; Gsh1 and Gsh2 [14]; Mox1 and Mox2 [15]; PBX1, PBX2 and PBX3 [16]; LFB1 and LFB3 [17]; several Pax genes; and many Oct genes. These proliferated genes seem to be located randomly at various chromosomal loci, based on the present knowledge [18,19].

In a previous study [20], we identified two new homeobox genes, GBX1 and GBX2, in the human genome by the PCR method. We characterized GBX1 and mapped it to chromosome 7q36.1 [20]. The member of another class of homeobox genes, EN-2, has also been mapped to this region [21]. Since both the EN-class and the GBX-class of homeobox genes have two members, we suspected that GBX2 might be mapped to a locus similar to that of EN-1, and that members of these two classes of homeobox genes could represent paralogous linkage groups.

2. MATERIALS AND METHODS

The GBX2 probe was described in a previous paper [20]. A human genomic library was screened with the GBX2 probe according to the Benton-Davis method [22]. Nucleotide sequence was determined by the dideoxy chain-termination method [23]. The details of the condi-

tions for fluorescence in situ hybridization (FISH) are described elsewhere [24,25]. A fragment corresponding to human EN-1 was amplified by PCR using two primers; 5'-GACAAGCGGCC(G/T)CG(G/C)AC(A/G)GC(G/C)TT(C/T)AC-3' and 5'-TGGTTGTACAG(T/G)-CCCTG(G/T)GCCATGAG-3'. A human genomic library was screened with this amplified fragment as a probe, and an EN-1-containing clone was isolated.

3. RESULTS

3.1. Structure of the human GBX2 gene

Preparation of the GBX2 probe was described in a previous paper [20]. A human genomic library was screened with the GBX2 probe. The nucleotide sequence of a homeobox-containing region was determined and indicated in Fig. 1a. Comparison of encoded amino acid sequences between GBX1 and GBX2 showed that the genes are closely related to one another (Fig. 1b) The degree of identity of amino acid residues in the homeobox, 58/60, is almost the same as that between EN-1 and EN-2 (57/60).

3.2. Mapping of the human GBX2 and EN-1 genes

The location of *GBX2* was determined by FISH. As shown in Fig. 2a, *GBX2* was mapped to chromosome 2q37. One of the HOX complexes, *HOX4D*, has been mapped to 2q31–37 [26]. In the mouse, this complex is located on chromosome 2 [27]. However, the mouse *En-1* gene is located on chromosome 1 [28], and the long arm of human chromosome 2 corresponds to mouse chromosomes 1 or 2, as indicated by comparative genome mapping [18,19]. In fact, human *EN-1* is located on chromosome 2 [21]. Very recently, *EN-1* was mapped to human chromosome 2q13-q21 using a mapping panel of rodent/human cell hybrids [29]. We examined the location of *EN-1* on human chromosome by FISH. As shown in Fig. 2b, *EN-1* was mapped to chro-

mosome 2q14, that supported the above observation [29]. Thus, we concluded that both *GBX1* and *EN-2* are located at chromosome 7q36 while both *GBX2* and *EN-1* are located on the same chromosome but at different loci, 2q37 and 2q14, respectively.

4. DISCUSSION

In the mouse, several alleles in which mutations cause abnormalities in skeletal development are known to be located in the vicinity of either En-1 or En-2 [30]. In the vicinity of En-1, there exists dominant hemimelia (Dh), a mutation that causes skeletal abnormalities in the hind limbs. In the vicinity of En-2, there exist two closely linked mutations, hemimelic extratoes (Hx) and hammertoe (Hm), which cause skeletal defects in all four limbs. Martin et al. [30] reported that the distance between the En-1 and Dh genes and that between the En-2 and Hx genes are 0.28 cM and 1.1 cM, respectively, and they proposed that En-1-Dh and En-2-Hx represent paralogous linkage groups that evolved after duplication of a common ancestral chromosome segment [30]. When we started the present experiment, we suspected that the ancestral segment may have contained members of both the EN-class and the GBX-class of homeobox genes. However, EN-1 and GBX-2 are located on the same chromosome but not closely linked to each other. One of the genes, in which a mutation causes defects of the development of brain and face, holoprosencephaly, has been mapped to human chromosome 7q36 [31].

It has been argued that *HOX*-containing regions on chromosomes 2, 7, 12 and 17 arose by regional duplication or by tetraploidization and that genes for collagen are contained in these paralogous chromosomal regions [32,33]. The location of *GBX2* seems to be close to that of the *HOXD* complex. Although, in mouse, the *HoxD*

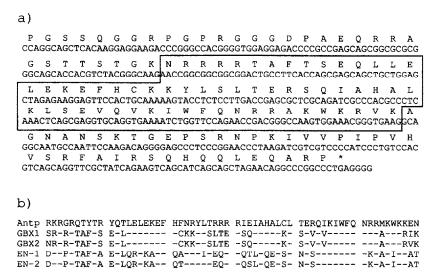


Fig. 1. Nucleotide and amino acid sequences of the human GBX2 gene. (a) Nucleotide and amino acid sequences of a homeobox-containing region of the GBX2 gene. The homeobox region is boxed. (b) Comparison of amino acid sequences of the homeoboxes. The amino acid sequence of Antp is used as a reference [37]. Each bar indicates an identical amino acid to that in Antp. Amino acid sequences of GBX1 and EN-1/EN-2 are taken from refs. [20] and [34], respectively.

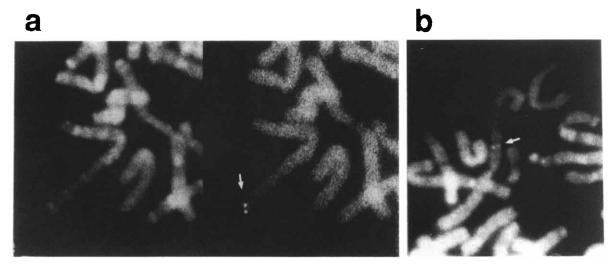


Fig. 2. Fluorescence in situ hybridization (FISH) of biotinylated human GBX2 and EN-1 probes on human chromosomes. (a) Mapping of GBX2 to chromosome 2q37. A 2.5-kb KpnI-AccI fragment containing GBX2 was used as the probe. Partial R-banded metaphase chromosome spread was viewed with a Nikon G-2A filter, showing R-bands (left). The same sample was viewed with a Nikon B-2E filter (right). An arrow indicates hybridization signals. (b) Mapping of EN-1 to chromosome 2q14. An 11.3-kb BamHI-EcoRI fragment containing EN-1 was used as the probe for FISH. Partial metaphase spread was viewed with a Nikon B-2A filter, showing R-bands and hybridization signals (arrow).

complex and En-1 are located on different chromosomes, the *Col6a-3* gene is mapped close to *En-1* [18,34]. Moreover, the COL6A3 gene is located at human chromosome 2q37 [34]. The EN-2/GBXI cluster is located on the same chromosome as HOXA in human. Thus, it is possible that the HOX complex, the EVX gene, a gene for collagen, the EN gene and the GBX gene were linked to each other on an ancestral genome. Although members of multiplied homeobox genes other than HOX genes seemed to be located randomly at various chromosomal loci, some of them may have been organized similarly to HOX, EVX, EN and GBX. Increases in the complexity of the body and organs in vertebrates may have been achieved by such multiplication, with subsequent diversification, of regions of DNA that contained genes for regulatory and structural proteins [32]. Possible clustering of members of different classes of homeobox genes has been suggested for Gsh-3, H6 and HOX7 at human chromosome 4q16 [35]. Just before we submit this paper, Komuro and Izumo [36] reported that two newly identified homeobox genes, Kbx and Imx, are closely linked to each other.

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REFERENCES

 McGinnis, W., Levine, M.S., Hafen, E., Kuroiwa, A. and Gehring, W.J. (1984) Nature 308, 428-433.

- [2] Scott, M.P., Tamkun, J.W. and Hartzell III, G.W. (1989) Biochem. Biophys. Acta 989, 25–48.
- [3] Colberg-Poley, A.M., Voss, S.D., Chowdhury, K. and Gruss, P. (1985) Nature 314, 713-718.
- [4] Singh, G., Kaur, S., Stock, J.L., Jenkins, N.A., Gilbert, D.J., Copeland, N.G. and Potter, S. (1991) Proc. Natl. Acad. Sci. USA 88, 10706-10710.
- [5] Murtha, M.T., Leckman, J.F. and Ruddle, F.H. (1991) Proc. Natl. Acad. Sci. USA 88, 10711-10715.
- [6] Kamps, M.P., Look, A.T. and Baltimore, D. (1991) Genes Dev. 5, 358-368.
- [7] Scott, M.P. (1992) Cell 71, 551-553.
- [8] Graham, A., Papalopulu, N. and Krumlauf, R. (1989) Cell 57, 367, 378
- [9] Faielle, A., D'Esposito, M., Rambaldi, M., Acampora, D., Balsofiore, S., Stornaiuolo, A., Mallamaci, A., Migliaccio, E., Gulisano, M., Simeone, A. and Boncinelli, E. (1991) Nucleic Acids Res. 19, 6541-6545.
- [10] D'Esposito, M., Morelli, F., Acampora, D., Migliaccio, E., Simeone, A. and Boncinelli, E. (1991) Genomics 10, 43-50.
- [11] Joyner, A.L. and Martin, G.R. (1987) Genes Dev. 1, 29-38.
- [12] German, M.S., Wang, J., Chadwick, R.B. and Rutter, W.J. (1992) Genes Dev. 6, 2165-2176.
- [13] Monaghan, A.P., Davidson, D.R.,, Sime, C., Graham, E., Baldock, R., Bhattacharya, S.S. and Hill, R.E. (1991) Development 112, 1053-1061.
- [14] Simeone, A., Gulisano, M., Acampora, D., Stornaiuolo, A., Rambaldi, M. and Boncinelli, E. (1992) EMBO J. 11, 2541-2550.
- [15] Candia, A.F., Hu, J., Crosby, J., Lalley, P.A., Noden, D., Nadeau, J.H. and Wright, C.V.E. (1992) Development 116, 1123– 1136.
- [16] Monica, K., Galili, N., Nourse, J., Saltman, D. and Cleary, M.L. (1991) Mol. Cell. Biol. 11, 6149–6157.
- [17] De Simone, V., De Magistris, L., Lazzaro, D., Gerstner, J., Monaci, P., Nicosia, A. and Cortese, R. (1991) EMBO J. 10, 1435–1443.
- [18] Nadeau, J.H., Davisson, M.T., Doolittle, D.P., Grant, P. Hillyard, A.L., Kosowsky, M.R. and Roderick, T.H. (1992) Mammal. Genome 3, 480-536.
- [19] O'Brien, S.J., Womack, J.E., Lyons, L.A., Moore, K.J., Jenkins, N.A. and Copeland, N.G. (1993) Nature Genet. 3, 103-112.

- [20] Matsui, T., Hirai, M., Wakita, M., Hirano, M. and Kurosawa, Y. (1993) FEBS Lett. 322, 181–185.
- [21] Logan, C., Willard, H.F., Rommens, J.M. and Joyner, A.L. (1989) Genomics 4, 206–209.
- [22] Benton, D.W. and Davis, R.W. (1977) Science 196, 180-182.
- [23] Sanger, F., Nicklen, S. and Coulson, A.R. (1977) Proc. Natl. Acad. Sci. USA 74, 5463-5467.
- [24] Takahashi, E., Hori, T., O'Connell, P., Leppert, M. and White, R. (1990) Hum. Genet. 86, 14-16.
- [25] Ichihara, Y., Hirai, M. and Kurosawa, Y. (1992) Immunol. Lett. 33, 277-284.
- [26] Cannizzaro, L.A., Croce, C.M., Griffin, C.A., Simeone, A., Boncinelli, E. and Huebner, K. (1987) Am. J. Hum. Genet. 41, 1-15.
- [27] Stubbs, L., Huxley, C., Hogan, B., Evans, T., Fried, M., Duboule, D. and Lehrach, H. (1990) Genomics 6, 645-650.
- [28] Hill, R.E., Jones, P.F., Rees, A.R., Sime, C.M. and Hastie, N.D. (1987) Cytogenet. Cell Genet. 44, 171-174.
- [29] Köhler, A., Logan, C., Joyner, A.L. and Muenke, M. (1993) Genomics 15, 233-235.

- [30] Martin, G.R., Richman, M., Reinsch, S., Nadeau, J.H. and Joyner, A. (1990) Genomics 6, 302-308.
- [31] Gurrieri, F., Trask, B.J., van den Engh, G., Krauss, C.M., Schinzel, A., Pettenati, M.J., Schindler, D., Dietz-Band, J., Vergnaud, G., Scherer, S.W., Tsui, L.-C. and Muenke, M. (1993) Nature Genet. 3, 247-251.
- [32] Ohno, S. (1970) in: Evolution by Gene Duplication, Springer, Berlin/Heidelberg/New York.
- [33] Lundin, L.G. (1993) Genomics 16, 1-19.
- [34] Schurr, E., Skamene, E., Morgan, K., Chu, M.-L. and Gros, P. (1990) Genomics 8, 477–486.
- [35] Stadler, H.S., Padanilam, B.J., Buetow, K., Murray, J.C. and Solursh, M. (1992) Proc. Natl. Acad. Sci. USA 89, 11579-11583.
- [36] Komuro, I. and Izumo, S. (1993) Proc. Natl. Acad. Sci. USA 90, 8145–8149.
- [37] Laughon, A., Boulet, A.M., Bermingham, Jr. J.R., Laymon, R.A. and Scott, M.P. (1986) Mol. Cell. Biol. 6, 4676-4689.